AD-A233 191





Transmission of Venezuelan Equine Encephalomyelitis
Virus by Strains of Aedes albopictus (Diptera: Culicidae)
Collected in North and South America

JOSEPH R. BEAMAN AND MICHAEL J. TURELL

Virology Division, U.S. Army Medical Research Institute of Infectious Diseases, Fort Detrick, Frederick, Maryland 21702 ELECTE MAR 2 9 1991

J. Med. Entomol. 28(1): 161-164 (1991)

ABSTRACT Experimental studies were undertaken to ascertain the vector potential of North American (Houston and Alsace) and South American (Sao Paulo and Santa Teresa) strains of Aedes albopictus (Skuse) for an epizootic (Trinidad donkey) strain of Venezuelan equine encephalomyelitis (VEE) virus. Infection rates were similar in all four strains of Aealbopictus tested after ingestion of VEE virus from a viremic hamster. Virus disseminated from the midgut to the hemocoel in about 80% of infected mosquitoes, regardless of the dose ingested (10⁴⁸ to 10⁵⁷ plaque-forming units per mosquito) or the time of extrinsic incubation (7–35 d). Although all four strains of this mosquito transmitted VEE virus by bite to hamsters, transmission rates were significantly higher for the South American strains (24%, 40 of 170) than for the North American strains (5%, 9 of 165). Although VEE virus has never been isolated from Ae. albopictus, the introduction of this species into the Americas may allow it to serve as an amplification vector in areas where epizootic strains of VEE are found or introduced.

KEY WORDS Insecta, *Aedes alboptctus*, Venezuelan equine encephalomyelitis virus, vector potentials

THE RECENT INTRODUCTION of Aedes albopictus (Skuse) into the Americas has raised a concern that these mosquitoes may serve as a vector for endemic as well as exotic viruses (Knudson 1986). Laboratory studies have demonstrated the ability of this species to transmit numerous arboviruses, including some native to the Americas (Shroyer 1986, Hawley 1988). In addition, Ae. albopictus can displace populations of Aedes aegypti (L.) in urban environments (Black et al. 1989) and has demonstrated an ability to flourish in arboreal settings (Rai 1986).

North and South American strains of Ae. albopictus differ from each other in several life history characteristics, including egg diapause, photoperiod sensitivity, and bloodmeal size (G. B. Craig, personal communication). Differences in vector competence for dengue viruses between North and South American strains also have been reported (Miller & Ballinger 1988). The strains of Ae. albopictus introduced into North America are presumed to have originated in temperate Asia, possibly Japan (Hawley et al. 1987). In contrast, strains

introduced into South America may have come from a tropical region of Asia (Hawley 1988).

Venezuelan equine encephalomyelitis (VEE) virus has caused sporadic epizootics of severe disease, usually fatal in horses and occasionally so in man, primarily in Central America (Walton & Grayson 1989). Epizootics have extended into northern South America and have occurred as far north as Texas in 1969–1972. Because of the potential for Ae. albopictus to transmit VEE virus, we evaluated two North American and two South American strains of Ae. albopictus for their ability to transmit an epizootic variant of VEE virus under laboratory conditions.

Materials and Methods

Mosquitoes. North American strains of Ae. albopictus were the Houston strain (Mitchell et al. 1987), derived from specimens collected in Houston, Tex., and obtained from Carl Mitchell, Centers for Disease Control, Ft. Collins, Colo.; and the Alsace strain, derived from specimens collected in Indiana and obtained from George Craig, Jr., University of Notre Dame, South Bend, Ind. (Table 1). South American strains were derived from specimens collected in Sao Paulo, Sao Paulo and Santa Teresa, Espirito Santo, Brazil (Sao Paulo and Santa Teresa, respectively). These strains also were obtained from Dr. Craig. Mosquitoes were maintained at 26°C using procedures described by Gargan et al. (1983), and

This document has been approved for public release and sale; its distribution is unlimited.

91 3 06 057

The views of the authors do not purport to reflect the positions of the Department of the Army or the Department of Defense. In conducting the research described in this report, the investigators adhered to the "Guide for the Care and Use of Laboratory Animals," as promulgated by the Committee on Care and Use of Laboratory Animals of the Institute of Laboratory Animal Resources, National Research Council. The facilities are fully accredited by the American Association for Accreditation of Laboratory Animal Care.

Table 1. Strains of Ac. albopictus tested in infection and transmission experiments

Strain	Collection location (year collected)	Generation
Houston	Houston, Tex. (1985)	F5-F9
Alsace	New Alsace, Ind. (1987)	F ₂ -F ₅
Santa Teresa	Santa Teresa, Espirito Santo, Brazil (1987)	F ₂ -F ₅
Sao Paulo	Sao Paulo, Sao Paulo, Brazil (1987)	F ₂ -F ₅

female mosquitoes were 4-10 d old when used for infection trials.

Virus and Virus Assay. A plaque-purified clone (FC-1-1) of the epizootic VEE subtype 1A Trinidad donkey strain (Johnston & Smith 1988) was used throughout these studies. It was passaged twice in BHK 21 cells and once in Vero cells after it had been cloned.

Serial 10-fold dilutions of specimens were plaqueassayed for infectious virus on Vero cell monolayers.

Determination of Vector Potential. Mosquitoes were allowed to feed on anesthetized female Syrian hamsters that had been inoculated intraperitoneally 96 h earlier with 0.2 ml of a suspension containing 10^{a5} plaque-forming units (PFU) of VEE virus. In most feedings, the anesthetized hamster was placed so that both a North American and a South American strain of Ae. albopictus could feed on it concurrently, thus insuring equal virus exposure to both strains. Immediately after feeding, five engorged mosquitoes from each hamster were triturated individually in 1 ml of diluent (10% fetal bovine serum in Medium 199 with Hanks' salts and antibiotics), frozen at -70°C, and then assayed on Vero cell monolayers to determine the amount of virus ingested. The remaining engorged mosquitoes were placed in a 3.8-liter cardboard container with netting on one end. Apple slices or a 7% sucrose solution was provided as a carbohydrate source. An oviposition substrate was added 4 d after the infectious blood meal. At 7-d intervals after the infectious blood meal, transmission attempts were made by allowing a sample of mosquitoes to feed. either individually or in pools of up to five mosquitoes each, on susceptible hamsters. Immediately after each transmission trial, mosquitoes were coldanesthetized, their legs and bodies triturated separately in 1 ml of diluent, and frozen at -70°C.

Infection was determined by the recovery of virus from the mosquito body tissue samples at ≥7 d after the infectious blood meal. A mosquito that had virus recovered from its body (but not its legs) was considered to have a nondisseminated infection limited to its midgut. In contrast, if virus was recovered from both body and leg suspensions, the mosquito was considered to have a disseminated infection (Turell et al. 1984). Because infection with VEE virus is virtually 100% fatal for hamsters, hamster death was used as the criterion for viral

Table 2. Infection and dissemination rates in selected strains of Ae. albopictus after ingestion of 10^{4.6}–10^{5.7} PFU of Venezuelan equine encephalomyelitis virus

Strain	No. tested	No. in- fected (%)	No. dis- seminated (%)
Houston	127	68 (54)	50 (39)
Alsace	91	67 (74)	53 (58)
Total, U.S. strains	218	135 (62)	103 (47)
Santa Teresa	189	133 (70)	110 (58)
Sao Paulo	168	104 (62)	84 (50)
Total, Brazilian strains	357	237 (66)	194 (54)

transmission. Isolation of virus from liver or brain tissue samples, or both, verified transmission. Any hamster that did not die after being fed upon by a mosquito with a disseminated infection was challenged with 10^{3.5} PFU of virus to confirm its susceptibility.

Results

Mosquitoes ingested 1046-1057 PFU of VEE virus in the various feedings. Within each strain, no consistent differences were observed in infection, dissemination, or transmission rates, regardless of the dose of VEE virus ingested. Thus, data for each strain were combined for further analysis. Infection and dissemination rates were similar in all of the Brazilian and North American strains tested (Table 2), but transmission rates differed (Tables 3 and 4). Transmission was first observed on day 7 after the infectious blood meal, and transmission rates were consistently higher at each time period for the Brazilian strains compared with the U.S. strains (Table 3). Transmission rates by Brazilian strains (24%, 40 of 170), were significantly higher $(\chi^2 = 20.5; df = 1; P < 0.001)$ than by U.S. strains (5%, 9 of 165), and this difference could be attributed to the percentage of mosquitoes with a disseminated infection that transmitted virus (Table

For both the North and South American strains, the virus titers recovered from mosquitoes with a disseminated infection were similar for transmitting (10^{5.7} and 10^{5.8} PFU per mosquito, respectively) and nontransmitting (10^{5.8} and 10^{5.7} PFU per mosquito, respectively) individuals. Also, the distribution of titers recovered from all individual infected mosquitoes was similar for the North and South American strains (Fig. 1).

Discussion

Strains of Ae. albopictus derived from specimens collected in both North and South America were competent vectors of VEE virus. Although both infection and dissemination rates, as well as the

ſ.

The second secon

Table 3. Transmission rates by day of extrinsic incubation in selected strains of As. albopictus after ingestion of $10^{4.6}$ – $10^{5.7}$ PFU of Venezuelan equine encephalomyelitis virus

Strain —	Day of extrinsic incubation				Totals	
	7	14	21	≥28	1 ocaus	
Houston Alsace	1/39 (3%) 0/41 (0%)	3/25 (12%) 3/25 (12%)	0/11 (0%) 1/6 (17%)	1/18 (6%)	5/93 (5%) 4/72 (6%)	
Total, U.S. strains	1/80 (1%)	6/50 (12%)	1/17 (6%)	1/18 (6%)	9/165 (5%)	
Santa Teresa Sao Paulo	3/22 (14%) 4/28 (14%)	6/27 (22%) 6/25 (24%)	5/11 (45%) 8/21 (38%)	5/18 (27%) 3/18 (17%)	19/78 (24%) 21/92 (23%)	
Total, Brazilian strains	7/50 (14%)	12/52 (23%)	13/32 (41%)	8/36 (22%)	40/170 (24%)	

Number of mosquitoes transmitting/number refeeding (percentage transmitting).

distribution of VEE viral titers recovered from individual mosquitoes, were similar, the South American strains were significantly more efficient laboratory vectors of VEE virus than were the North American strains. This is in contrast to the greater susceptibility to all four serotypes of dengue viruses by the Houston strain of Ae. albopictus compared with a Brazilian strain of Ae. albopictus (Miller & Ballinger 1988).

The difference in transmission rates between the North and South American strains of Ae. albopictus in the present study probably is due to either a salivary gland infection or a salivary gland escape barrier (Kramer et al. 1981) because only 16% of the females of the U.S. strains with a disseminated infection transmitted VEE virus compared with 60% of the disseminated refeeding Brazilian mosquitoes. Interestingly, for mosquitoes with a disseminated infection, there were no significant differences in titers recovered from a mosquito, regardless of its transmission status (transmitter or nontransmitter) or its origin (North or South America).

The transmission rate we observed for the South American strains (24%, 40 of 170), is similar to that reported for *Psorophora confinnis* (Lynch Arribalzaga) (33%, 6 of 18) (Sudia et al. 1971b). This species has been implicated as a natural vector of epizootic VEE virus (Sudia et al. 1971a) and, of all

Table 4. Transmission rates in selected strains of Ae. albopictus after ingestion of 10^{4.6}–10^{5.7} PFU of Venezuelan equine encephalomyelitis virus

Strain	All refeedings	Disseminated, refeeding ^b	
Houston	5/93 (5%)	5/26 (19%)	
Alsace	4/72 (6%)	4/31 (13%)	
Total, U.S. strains	9/165 (5%)	9/57 (16%)	
Santa Teresa	19/78 (24%)	19/33 (58%)	
Sao Paulo	21/92 (23%)	21/33 (64%)	
Total, Brazilian strains	40/170 (24%)	40/66 (61%)	

Number of mosquitoes transmitting/number refeeding (percentage transmitting).

the mosquito species tested, was the most efficient transmitter for an epizootic strain of VEE virus (Sudia et al. 1971b).

The hamster viremias to which mosquitoes were exposed in the present study are comparable with those observed in burros (Gochenour et al. 1962) and in horses (Kissling et al. 1956, Sudia et al. 1971b) inoculated with an epizootic strain of VEE virus. Ae. albopictus readily feeds on humans, as evidenced by its role in the transmission of dengue virus (Metselaar et al. 1980). In addition, it avidly feeds on many other mammals, including horses (Tempelis et al. 1970). This mosquito has extended its range in many areas, including areas where VEE has been epizootic in North America. Thus, Ae. albopictus, especially the Brazilian strains, should be considered a potential vector of epizootic VEE virus.

Acknowledgment

We thank J. Kondig and his insectary staff for their assistance in rearing the mosquitoes, and D. Dickson and S. Gordon for their critical reading of the manuscript.

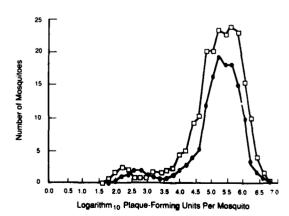


Fig. 1. Distribution of viral titers recovered from North and South American strains of Ae. albopictus tested 7-35 d after ingestion of VEE virus.

On North American strains.

^b Number of mosquitoes transmitting/number with a disseminated infection refeeding (percentage transmitting).

References Cited

- Black, W. C. IV, K. S. Rai, B. J. Turce & P. C. Arroyo. 1989. Laboratory study of competition between United States strains of Aedes albopictus and Aedes aegypti (Diptera: Culicidae). J. Med. Entomol. 26: 280-271.
- Gargan, T. P., II, C. L. Bailey, G. A. Higbee, A. Gad & S. El Said. 1983. The effect of laboratory colonization on the vector pathogen interactions of Egyptian *Culex pipiens* and Rift Valley fever virus. Am. J. Trop. Med. Hyg. 32: 1154-1163.
- Gochenour, W. S., Jr., T. O. Berge, C. A. Gleiser & W. D. Tigertt. 1962. Immunization of burros with living Venezuelan equine encephalomyelitis virus. Am. J. Hyg. 75: 351–362.
- Hawley, W. A. 1988. The biology of Aedes albopictus. J. Am. Mosq. Control Assoc. (Suppl.) 4: 1-39.
- Hawley, W. A., P. Reiter, R. S. Copeland, C. B. Pumpuni & G. B. Craig, Jr. 1987. Aedes albopictus in North America: probable introduction in used tires from northern Asia. Science 236: 1114–1116.
- Johnston, R. E. & J. F. Smith. 1988. Selection for accelerated penetration in cell culture coselects for attenuated mutants of Venezuelan equine encephalitis virus. Virology 162: 437-443.
- Kissling, R. E., R. W. Chamberlain, D. B. Nelson & D. D. Stamm. 1956. Venezuelan equine encephalomyelitis in horses. Am. J. Hyg. 63: 274–287.
- Knudson, A. B. 1986. The significance of the introduction of Aedes albopictus into the southeastern United States with implications for the Caribbean and perspectives of the Pan American Health Organization. J. Am. Mosq. Control Assoc. 2: 420-423.
- Kramer, L. D., J. L. Hardy, S. B. Presser & E. J. Houk. 198 Dissemination barriers for western equine encephalomyelitis virus in *Culex tarsalis* infected after ingestion of low viral doses. Am. J. Trop. Med. Hyg. 30: 190-197.
- Metselaar, D., C. R. Grainger, K. G. Oei, D. G. Reynolda, M. Pudney, C. J. Leake, R. M. Tukei, R. M. D'Offay & D.I.H. Simpson. 1980. An outbreak

- of type 2 dengue fever in the Seychelles probably transmitted by Aedes albopictus (Skuse). Bull. W. H. O. 58: 937-943.
- Miller, B. R. & M. E. Ballinger. 1988. Aedes albopictus mosquitoes introduced into Brazil: vector competence for yellow fever and dengue viruses. Trans. R. Soc. Trop. Med. Hyg. 82: 476-477.
- Mitchell, C. J., B. R. Miller & D. J. Gubler. 1987.

 Vector competence of *Aedes albopictus* from Houston, Texas, for dengue serotypes 1 to 4, yellow fever and Ross River viruses. J. Am. Mosq. Control Assoc. 3: 460-465.
- Rai, K. S. 1986. Genetics of Aedes albopictus. J. Am. Mosq. Control Assoc. 2: 429–436.
- Shroyer, D. A. 1986. Aedes albopictus and arboviruses: a concise review of the literature. J. Am. Mosq. Control Assoc. 4: 424-428.
- Sudia, W. D., R. D. Lord, V. F. Newhouse, D. L. Miller & R. E. Kissling. 1971a. Vector-host studies of an epizootic of Venezuelan equine encephalomyelitis in Guatemala, 1969. Am. J. Epidemiol. 93: 137-143.
- Sudia, W. D., V. F. Newhouse & B. E. Henderson. 1971b. Experimental infection of horses with three strains of Venezuelan equine encephalomyelitis II. Experimental vector studies. Am. J. Epidemiol. 93: 206-211.
- Turell, M. J., T. P. Gargan II, & C. L. Bailey. 1984. Replication and dissemination of Rift Valley fever virus in *Culex pipiens*. Am. J. Trop. Med. Hyg. 33: 176-181.
- Tempelia, C. H., R. O. Hayes, A. D. Hess & W. C. Reeves. 1970. Blood feeding habits of four species of mosquito found in Hawaii. Am. J. Trop. Med. Hyg. 19: 335-341.
- Walton, T. E. & M. A. Grayson. 1989. Venezuelan equine encephalomyelitis, pp. 203–231. In T. P. Monath [ed.], The arboviruses: epidemiology and ecology, vol. 4, CRC, Boca Raton, Fla.

Received for publication 22 February 1990; accepted 17 July 1990.

Accesi	Accesion For			
DTIC Unann	NTIS CRA&I DTIC TAB Unannounced Justification			
By				
Availability Codes				
Dist	Avail and/or Special			
AI	20			

